



Policy Forums

Management of vampire bats and rabies: a precaution for rewilding projects in the Neotropics



Fernando Gonçalves^{a,b,*}, Mauro Galetti^c, Daniel G. Streicker^{d,e}

^a Institute of Bioscience, Department of Biodiversity, Universidade Estadual Paulista (UNESP), Rio Claro, Brazil

^b Conservation Science Group, Department of Zoology, University of Cambridge, Cambridge, UK

^c Department of Biology, University of Miami, Coral Gables, FL, USA

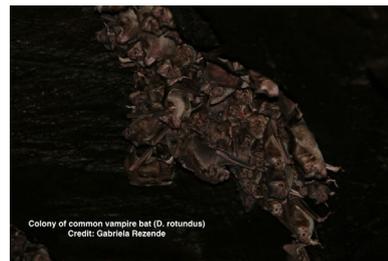
^d Institute of Biodiversity, Animal Health and Comparative Medicine, College of Medical, Veterinary and Life Sciences, University of Glasgow, Glasgow, UK

^e MRC–University of Glasgow Centre for Virus Research, Glasgow, UK

HIGHLIGHTS

- Certainly attention needs to be brought to the potential consequences of creating novel animal communities on disease transmission
- Bats host pathogens that cause significant human and animal morbidity and mortality.
- We highlight how to prevent, detect and mitigate vampire bats and rabies in rewilding projects.

GRAPHICAL ABSTRACT



ARTICLE INFO

Article history:

Received 2 September 2020

Accepted 11 December 2020

Available online 10 February 2021

Keywords:

Bat-transmitted pathogen
Multidisciplinary teams
Animal restoration
Oral vaccines
Reproductive control
Human and animal mortality

ABSTRACT

Changes in animal population dynamics and community composition following species (re)introduction may have unanticipated consequences for a variety of downstream ecosystem processes, including infectious disease transmission. Due the lessons learned from ongoing projects, we present a novel approach on how to anticipate, monitor, and mitigate the vampire bats and rabies in rewilding projects. We pinpoint a series of precautions and the need for long-term monitoring of vampire bats and rabies responses to rewilding projects and highlighted the importance of multidisciplinary teams of scientist and managers focusing on prevention educational program of rabies risk transmitted by bats. In addition, monitoring the relative abundance of vampire bats, considering reproductive control by sterilization and oral vaccines that autonomously transfer among bats would reduce the probability, size and duration of rabies outbreaks. The rewilding assessment framework presented here responds to calls to better integrate the science and practice of rewilding and will help conservation practitioners and researchers to develop effective message framing strategies that minimize bats emerging infectious diseases and support biodiversity and its associated ecosystem services.

© 2021 Associação Brasileira de Ciência Ecológica e Conservação. Published by Elsevier Editora Ltda.

This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Introduction

The urgent need to reverse anthropogenic impacts on ecosystems and biodiversity is one of the biggest challenges for modern society (Svenning et al., 2016; Perino et al., 2019; Moreno-Mateos

* Corresponding author.

E-mail address: fhm.goncalves@unesp.br (F. Gonçalves).

Table 1
Prey species observed bitten by vampire bats in the Neotropical area.

Prey	Common name	Country	Bioregion	Ref
Reptile				
<i>Elaphe flavirufa</i>	Yellow-red Rat Snake	Mexico	Mesoamerica	1
Mammals				
<i>Callicebus aureipalatii</i>	Madidi Titi Monkey	Peru	Amazon	2
Coendu sp.	Porcupine	Mexico	Mesoamerica	3
<i>Dasyus</i> sp.	armadillo	Mexico	Mesoamerica	3
<i>Hydrochaeris hydrochaeris</i>	Capybara	Brazil, Venezuela	Atlantic forest, Illanos	4,5,6,7
<i>Mazama americana</i>	Red Brocket	Brazil	Atlantic forest, Pantanal	8,9
<i>Odocoileus virginianus</i>	White-tailed Deer	Mexico	Mesoamerica	10
<i>Otaria flavescens</i>	Southern Sea Lion	Peru, Chile	Coast, Island	2,11,12
<i>Pecari tajacu</i>	Collared Peccary	Brazil	Atlantic forest	8
<i>Priodontes maximus</i>	Giant Armadillo	Brazil	Atlantic forest	8
<i>Saimiri boliviensis</i>	Squirrel Monkey	Peru	Amazon	2
<i>Sturnira lilium</i> ^a	Yellow-shouldered Bat	Argentina	Dry forest	13
<i>Sus scrofa</i>	Wild Boar	Brazil, Mexico	Atlantic forest, Pantanal, Mesoamerica	9,14
<i>Tapirus terrestris</i>	Lowland Tapir	Peru, Brazil	Amazon, Atlantic forest, Pantanal	2,8,9
<i>Tayassu pecari</i>	White-lipped Peccary	Peru	Amazon	2
<i>Sciurus</i> sp.	Squirrel	Mexico	Mesoamerica	16
Birds				
Cormorants	sea birds	Chile	Island	12
Pelicans	sea birds	Chile	Island	12
<i>Spheniscus humboldti</i>	Humboldt Penguin	Chile	Island	17

1-Villa and Lopez-Forment, 1966; 2-Streicker and Allgeier, 2016; 3-Greenhall et al., 1983; 4-Gonçalves et al., 2020; 5-Azcarate, 1980; 6-Ibanez, 1981; 7-Carranza, 1982; 8-Zortéa et al., 2018; 9-Galetti et al., 2016; 10-Sánchez-Cordero et al., 2011; 11-Catenazzi and Donnelly, 2008; 12-Mann, 1951; 13-Lord et al., 1973; 14-Hernández-Pérez et al., 2019; 15-Gnocchi and Srbeć-Araujo, 2017; 16-Greenhall, 1972; 17-Luna-Jorquera and Culiik, 1995.

^a Captured carried distinct vampire bats wounds.

et al., 2020). Discussions on post-2020 biodiversity strategies by the signatory countries of the Convention on Biological Diversity are currently being initiated, and the United Nations General Assembly has recently declared 2021–2030 the “decade of ecosystem restoration” (United Nations, 2019). Due the designation, policy- and decision-makers will push animal restoration topics to the forefront of discussions about how to reach post-2021 biodiversity goals, especially because restoration projects could provide a buffer to species extinction (Galetti et al., 2017) and restore plant–animal interactions and ecological processes impaired by defaunation (Pires, 2017; Genes et al., 2019; Mittelman et al., 2020). However, changes in animal population dynamics and community composition following species (re)introduction may have unanticipated consequences for a variety of downstream ecosystem processes, including food web structure (Lovari et al., 2014), predator–prey systems (Bovendorp and Galetti, 2007) and infectious disease emergence (Lafferty and Gerber, 2002). This highlights the need to develop frameworks to anticipate, monitor, and mitigate the unintended ecosystem consequences of ‘re-wilding’ projects.

In recent years, bats have received growing attention as reservoirs of pathogens linked to emerging zoonotic diseases that cause significant human and animal morbidity and mortality. Key examples include Ebola, Nipah and potentially the ongoing COVID-19 pandemic, given that viral strains closely related to SARS-CoV-2 have been detected in bats (Letko et al., 2020). In the Neotropics, the most notorious bat-transmitted pathogen is rabies virus (Rhabdoviridae, Lyssavirus), which causes a nearly universally lethal infection in all mammals (including humans) and is maintained by a wide variety of bat species (Streicker et al., 2010; Fisher et al., 2018). Among the three species of vampire bats (*Desmodus rotundus*, *Diaemus youngi*, and *Diphylla ecaudata*) that rely on blood as their food source, *D. rotundus* is the only known reservoir for rabies. *D. rotundus* frequently bites domestic and wild animals and more rarely humans during feeding (Bobrowiec et al., 2015; Galetti et al., 2016; Gnocchi and Srbeć-Araujo, 2017; Zórtea et al., 2018; Gonçalves et al., 2020), creating opportunities for the transmission of rabies virus and potentially other saliva-borne viruses (Bergner et al., 2020).

Official guidelines for managing rabies transmitted by vampire bats involve pre-exposure vaccination of livestock, post-exposure vaccination of humans bitten by vampire bats (and more rarely pre-exposure human vaccination) and bat population control (Benavides et al., 2020a; Recuenco, 2020). Although bat culls sometimes involve vigilante action such as burning or destruction of bat roosts, governments actively discourage these non-specific actions in favor of topical, anticoagulant poisons (“vampiricides”) which spread intra-specifically by vampire bat grooming. Similar poisons can be applied to the wounds of bitten livestock, and ingested when bats return to feed (Thompson et al., 1972). Policies for vampire bat monitoring and initiating culls are country-specific and implemented by Ministries of Health and/or Ministries of Agriculture. In general, culls are reactive to reports of increased vampire bat bites on livestock or detection of rabies cases via passive surveillance systems. While culling effectively reduces vampire bat bites, some studies suggest that significant mitigation of the burden of rabies in humans and domestic livestock would require implementing culls over impractically large geographical areas (Streicker et al., 2012; Blackwood et al., 2013; Bakker et al., 2019). Other proposed strategies for controlling vampire bat populations involve modified farming practices that reduce the accessibility to bats (e.g., artificial lighting, protected corrals, altering the composition and locations of herds) or hormonal reproductive control of bats, though the latter is in early research stages (Benavides et al., 2020a).

D. rotundus has specialized on blood of medium to large-bodied mammal species and demonstrates strong plasticity of foraging strategies due to changes in local prey availability across regions (Table 1). Due to the proliferation of livestock production in much of the Neotropics and its relative reliability compared to native wildlife, livestock are generally preferred prey (Voigt and Kelm, 2006; Gonçalves et al., 2017; Bohmann et al., 2018). However, dietary shifts to humans and wildlife have also been reported following changes in local prey abundance. For example, in Brazil, conversion of pasture into plantation agriculture forced vampire bats switch from the formerly abundant livestock animals to wild animals (Galetti et al., 2016). In Belize, removal of livestock increased *D. rotundus* feeding on human beings (McCarthy, 1989) and removal of wildlife by hunting was speculated to increase

human rabies risk in Peru (Stoner-Duncan et al., 2014). In the context of rewilding, released species may be at particular risk since candidate areas for restoration tend to have low availability of medium to large-bodied wild prey due to historical human disturbance (Galetti et al., 2017; Fernandez et al., 2017). In addition, rewilding projects typically rely on captive born animals (Fernandez et al., 2017) that have diminished functional traits associated with adaptation to captivity, including exposure to predators (Jule et al., 2008), which could heighten vulnerability to vampire bats.

Recently, an unprecedented study in Brazil showed changes in vampire bat feeding following a rewilding project (Gonçalves et al., 2020). Specifically, in a land-bridge island where 100 individuals of 15 non-volant mammal species were introduced with the intent of “restoring” the local fauna, *D. rotundus* fed primarily on introduced capybaras (Gonçalves et al., 2020). Thirty-six years following restoration, the relative abundance of common vampire bats was higher on the islands than the mainland nearby due to the increased prey availability. The restoration further transformed the land-bridge island into a high-risk area for rabies transmission as three introduced capybaras were confirmed to have died from bat transmitted rabies (Gonçalves et al., 2020).

Due the lessons learned from ongoing rewilding projects combined with public health risk if rabies infections from (re)introduced species lead to human exposures, we present a novel approach on how to prevent and control vampire bats and rabies in rewilding projects (Fig. 1). While we highlight vampire bats as a focal example, we seek to establish a generalizable rewilding-monitoring framework that can anticipate and detect how indirect processes may alter the outcomes of restoration projects.

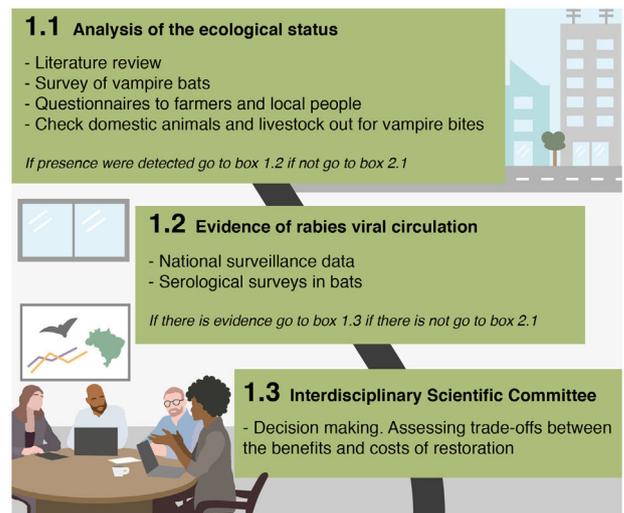
Phase 1: pre-risk assessment

The first step of the framework should assess the presence and infection status of natural reservoir hosts in the focal area. In the case of vampire bats, presence could be verified by field surveys or questionnaires to local people about bat bites on domestic animals (a strong indicator of *D. rotundus* presence). If vampire bats are confirmed present, viral circulation could be assessed using serological surveys in wild bats to detect antibodies from recent exposures (Meza et al., 2020). Since naïve domestic animals are increasingly recognized to sometimes survive and mount detectable antibody responses to rabies virus exposures, serological surveys of livestock reported to be un-vaccinated by their owners could similarly indicate viral circulation (Benavides et al., 2020b). Finally, recent records of rabies incidence in humans, domestic or wild animals from national surveillance systems could provide further evidence of rabies circulation. If vampire bats are present and there is evidence of viral circulation, multidisciplinary teams of scientists and managers should develop plans that assess the benefits and costs of the rewilding project, both in terms of the success of restoration (particularly how added rabies mortality would affect the initial establishment of rare/threatened species) and potential public health risks arising from human exposures to rabies cases in restored wildlife.

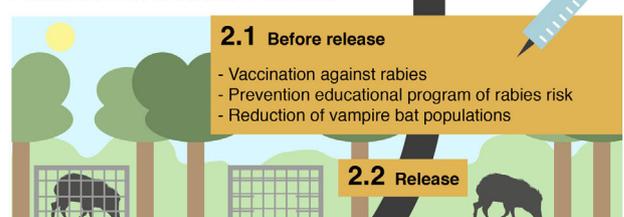
Phase 2: pre-release actions

Reduction of vampire bat populations may be considered prior to release by a multidisciplinary teams of scientist that have to evaluate the ethical issues on both, to mitigate effects on re-introduced prey (blood loss, rabies transmission) and on other bat species which may compete with *D. rotundus* for roost space. In much of its range, *D. rotundus* populations are controlled using topical, anti-coagulant poisons, which are either applied directly to captured

PHASE 1: PRE-RISK ASSESSMENT



PHASE 2: PRE-RELEASE ACTIONS



PHASE 3: POST-RELEASE

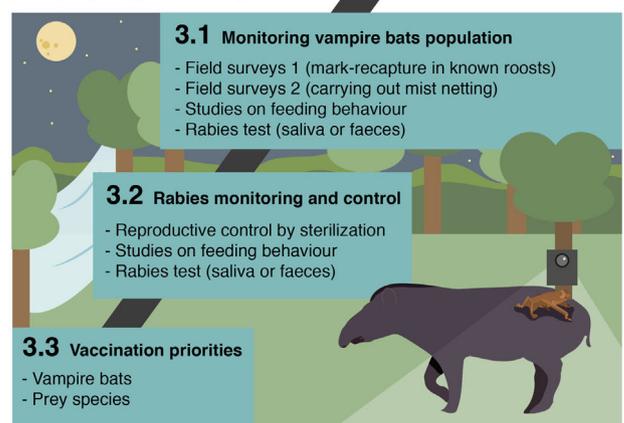


Fig. 1. Generalizable rewilding-monitoring framework that can anticipate and mitigate the unintended ecosystem consequences of ‘re-wilding’ projects.

bats and spread by grooming or can be applied to livestock to be ingested when bats feed (Johnson et al., 2014). Although social disruption of culls is predicted to increase rabies transmission in endemic areas (Streicker et al., 2012; Blackwood et al., 2013), culls carried out in rabies free populations may reduce the likelihood of viral incursions (Bakker et al., 2019). Reproductive control by sterilization (Serrano et al., 2007) is less explored alternative. Although the time lag between sterilization campaigns and diminished population sizes makes this tool ineffective at the pre-release stage, it could be a valuable long-term strategy to manage vampire bat population sizes (see phase 3).

Presumably, if vampires are present and rabies is detected, all mammals (> 1 kg) should be vaccinated against rabies before release. Under this phase, prevention educational program of rabies risk transmitted by bats should begin by a multi-disciplinary task force, included public and animal health services and universities,

focusing on local people living in the destined site and those actually working on the rewilding project.

Phase 3: post-release

Management of vampire bat populations

Monitoring presence and/or abundance of vampire bat is particularly challenging in the context of rewilding. Surveys of bat bites are unlikely to be appropriate since livestock will no longer be abundant following most habitat restoration projects and analogous monitoring of more elusive wildlife is not practical. Another indirect technique, acoustic monitoring, has limited utility due to the ‘soft’ echolocation of vampire bats (Rodríguez-San Pedro and Allendes, 2017). Consequently, labor-intensive field surveys may be the only option to monitor increases in vampire bat populations following restoration. Past studies have measured relative abundance using mark-recapture in known roosts or by carrying out mist netting away from roosts and potential prey sources (Delpietro et al., 1992; Streicker et al., 2012). In conjunction with capture efforts, studies on feeding behaviour [e.g. genetic analysis of blood meals (Bohmann et al., 2018) feces (Bobrowiec et al., 2015), camera trap (Galetti et al., 2016; Zortéa et al., 2018) and stable isotopes (Streicker and Allgeier, 2016; Gonçalves et al., 2020)] could monitor if introduced species (and which) are being exploited as a blood source in order to guide longer term vaccination priorities.

Rabies monitoring and control

Monitoring for rabies in the bats themselves can indicate viral circulation, but has limitations from a long term management perspective. The most commonly used antibody assays of rabies exposure necessarily lag behind transmission. Moreover, other bats (e.g. insectivores) may also have rabies and could cause seroconversion in *D. rotundus*; necessitating caution in interpretation of serological data. Monitoring by killing bats and testing their brains for rabies antigen is also unlikely to be productive since the prevalence of rabies is usually so low in randomly sampled bats (<1%). PCR based testing in saliva or feces might be considered, especially if field-based testing becomes available since several teams have shown that RNA can be detectable more readily than previously expected (Bergner et al., 2020; Begeman et al., 2020), but cost limits this approach to occasional evaluations rather than long term monitoring.

Since the long term persistence of rabies seems to be a spatial process (e.g., waves of infection that move across the landscape or metapopulation dynamics), cases in livestock or humans reported through existing national surveillance systems in areas near rewilding projects could function as an early warning system. Since rabies is an OIE notifiable disease, such surveillance systems are well established and routinely used, providing a wealth of data on the distribution of rabies. Moreover, genetic or antigenic typing of rabies positive animals could provide clear evidence of vampire bat rabies (as opposed to variants associated with other bats or terrestrial wildlife) circulation in or in close proximity to restoration projects (Fooks and Jackson, 2020). In most countries, analogous rabies diagnostic testing of suspicious wildlife is not routinely carried out in the absence of human exposure. Such testing might have greater priority in the context of restoration projects since restoration actions themselves are likely to reduce the presence of livestock available for rabies detection via existing passive surveillance systems. Whether relying on reports from domesticated or wild animals, communication between restoration managers and the epidemiologists and laboratory diagnosticians in rabies control programs will be vital.

Rabies circulation in vampire bats might also be incidentally detected through reports of unusual bat activity (e.g., daytime flight or discovery of moribund or dead bats) by the local public. Educational programs in the pre-release phase might emphasize the importance of reporting such events, to whom reports should be directed, and instructions for safe handling of moribund bats. Educational materials should convey not only the risk of human rabies from bats but also conservation messages about the broad ecological benefits of bats (e.g., seed dispersal, pollination).

If evidence of rabies circulation is detected, interventions could either target restored wildlife or the bats themselves. Non-bat species could be considered for vaccination based on observations of vampire bat bites or analysis of vampire bat diets in conjunction with an evaluation of the species-specific impacts of rabies losses for rewilding aims. Vaccination of bats may be effective, particularly given tentative evidence that vaccines that administered to bats that are already infected may stop them from transmitting (Cárdenas-Canales et al., 2020). The new and promising bat-to-bat transfer and ingestion oral vaccine (Bakker et al., 2019) may reduce costs and increase population level coverage. The vaccine is transferred among individuals through contacts due high rates of social grooming and, even at low but achievable levels of vaccine application, can control vampire bat rabies and could be applied either before or during outbreaks (Bakker et al., 2019).

Conclusion

We pinpoint a series of precautions and the need for long-term monitoring of vampire bats and rabies responses to rewilding projects. We highlighted the importance of multidisciplinary teams of scientist and managers, including national surveillance systems, focusing on prevention educational program of rabies risk transmitted by bats in rewilding projects. In addition, monitoring the relative abundance of vampire bats, considering reproductive control by sterilization and oral vaccines that autonomously transfer among bats would reduce the probability, size and duration of rabies outbreaks. The rewilding assessment framework presented here responds to calls to better integrate the science and practice of rewilding (Nogués-Bravo et al., 2016) and also the urgent need for long-term studying of bat-transmitted pathogen in the Neotropical area as the region is considered a geographic hotspots of “missing bat zoonoses” (Olival et al., 2017). Although there are challenges remaining, we believe that the implementation and further development of our monitoring framework will help catalyse a positive and ambitious vision for rewilding. Furthermore, the application of this framework provides guidance for practitioners, funders and decision-makers to incorporate or demand a multifaceted perspective for rewilding and, simultaneously, incentivize conservation initiatives to go beyond the recovery of species and habitats and include ecosystem function and processes. The framework is applicable to management a variety of bat-transmitted pathogens in restoration projects that aims to promote beneficial interactions between society and nature, range from conservation translocation to all variation of rewilding (Corlett, 2016).

Conflict of interest

The authors declare that there is no conflict of interest.

Acknowledgements

F.G. and M.G. was supported by (FAPESP) São Paulo Research Foundation (Grant 2017/24252-0, 2019/00648-7). D.G.S. was supported by a Wellcome Trust Senior Research Fellowship (Grant: 217221/Z/19/Z).

References

- Azcarate, T., 1980. *Sociobiología y manejo del capibara (Hydrochoerus hydrochaeris)*. *Acta Vert.* 7, 1–228.
- Bakker, K.M., Rocke, T.E., Osorio, J.E., Abbott, R.C., Tello, C., Carrera, J.E., Valderrama, W., Shiva, C., Falcon, N., Streicker, D.G., 2019. Fluorescent biomarkers demonstrate prospects for spreadable vaccines to control disease transmission in wild bats. *Nat. Ecol. Evol.* 3, 1697–1704. <http://dx.doi.org/10.1038/s41559-019-1032-x>.
- Begeman, L., Kooi, E.A., van Weezep, E., van de Bildt, M.W.G., Reusken, C.B.E.M., Lina, P.H.C., Koopmans, M.P.G., van den Brand, J.M.A., Kuiken, T., 2020. Faeces as a novel material to estimate lyssavirus prevalence in bat populations. *Zoonoses Public Health* 67, 198–202. <http://dx.doi.org/10.1111/zph.12672>.
- Benavides, J.A., Valderrama, W., Recuenco, S., Uieda, W., Suzán, G., Avila-Flores, R., Velasco-Villa, A., Almeida, M., de Andrade, F.A.G., Molina-Flores, B., Vigilato, M.A.N., Pompei, J.C.A., Tizzani, P., Carrera, J.E., Ibanez, D., Streicker, D.G., 2020a. Defining new pathways to manage the ongoing emergence of bat rabies in Latin America. *Viruses* 12. <http://dx.doi.org/10.3390/v12091002>.
- Benavides, J.A., Velasco-Villa, A., Godino, L.C., Satheshkumar, P.S., Ruby, N., Rojas-Paniagua, E., Shiva, C., Falcon, N., Streicker, D.G., 2020b. Abortive vampire bat rabies infections in peruvian peridomestic livestock. *PLoS Negl. Trop. Dis.* 14, 1–13. <http://dx.doi.org/10.1371/journal.pntd.0008194>.
- Bergner, L.M., Orton, R.J., Benavides, J.A., Becker, D.J., Tello, C., Biek, R., Streicker, D.G., 2020. Demographic and environmental drivers of metagenomic viral diversity in vampire bats. *Mol. Ecol.* 29, 26–39. <http://dx.doi.org/10.1111/mec.15250>.
- Blackwood, J.C., Streicker, D.G., Altizer, S., Rohani, P., 2013. Resolving the roles of immunity, pathogenesis, and immigration for rabies persistence in vampire bats. *Proc. Natl. Acad. Sci.* 110, 20837–20842. <http://dx.doi.org/10.1073/pnas.1308817110>.
- Bobrowiec, P.E.D., Lemes, M.R., Gribel, R., 2015. Prey preference of the common vampire bat (*Desmodus rotundus*, Chiroptera) using molecular analysis. *J. Mammal.* 96, 54–63. <http://dx.doi.org/10.1093/jmammal/gyu002>.
- Bohmann, K., Gopalakrishnan, S., Nielsen, M., Nielsen, Ldos S.B., Jones, G., Streicker, D.G., Gilbert, M.T.P., 2018. Using DNA metabarcoding for simultaneous inference of common vampire bat diet and population structure. *Mol. Ecol. Resour.* 18, 1050–1063. <http://dx.doi.org/10.1111/1755-0998.12891>.
- Bovendorp, R.S., Galetti, M., 2007. Density and population size of mammals introduced on a land-bridge island in southeastern Brazil. *Biol. Invasions* 9, 353–357. <http://dx.doi.org/10.1007/s10530-006-9031-7>.
- Cárdenas-Canales, E.M., Gigante, C.M., Greenberg, L., Velasco-Villa, A., Ellison, J.A., Satheshkumar, P.S., Medina-Magües, L.G., Griesser, R., Falendysz, E., Amezcua, I., Osorio, J.E., Rocke, T.E., 2020. Clinical presentation and serologic response during a rabies epizootic in captive common vampire bats (*desmodus rotundus*). *Trop. Med. Infect. Dis.* 5, 34. <http://dx.doi.org/10.3390/tropicalmed5010034>.
- Carranza, J., 1982. Murciélago hematófago *Desmodus rotundus* parasitando a un chiguire *Hydrochoerus hydrochaeris*. *Acta Vertebrata* 9, 414.
- Catenazzi, A., Donnelly, M.A., 2008. Sea lion *Otaria flavescens* as host of the common vampire bat *Desmodus rotundus*. *Mar. Ecol. Prog. Ser.* 360, 285–289.
- Corlett, R.T., 2016. Restoration, reintroduction, and rewilding in a changing world. *Trends Ecol. Evol. (Amst.)* 31, 453–462. <http://dx.doi.org/10.1016/j.tree.2016.02.017>.
- Delpietro, H.A., Marchevsky, N., Simonetti, E., 1992. Relative population densities and predation of the common vampire bat (*Desmodus rotundus*) in natural and cattle-raising areas in north-east Argentina. *Prev. Vet. Med.* 14, 13–20. [http://dx.doi.org/10.1016/0167-5877\(92\)90080-Y](http://dx.doi.org/10.1016/0167-5877(92)90080-Y).
- Fernandez, F.A.S., Rheingantz, M.L., Genes, L., Kenup, C.F., Galliez, M., Cezimbra, T., Cid, B., Macedo, L., Araujo, B.B.A., Moraes, B.S., Monjeau, A., Pires, A.S., 2017. Rewilding the Atlantic Forest: restoring the fauna and ecological interactions of a protected area. *Perspect. Ecol. Conserv.* 15, 308–314. <http://dx.doi.org/10.1016/j.pecon.2017.09.004>.
- Fisher, C.R., Streicker, D.G., Schnell, M.J., 2018. The spread and evolution of rabies virus: conquering new frontiers. *Nat. Rev. Microbiol.* 16, 241–255. <http://dx.doi.org/10.1038/nrmicro.2018.111>.
- Fooks, A.R., Jackson, A.C., 2020. *Rabies: Scientific Basis of the Disease and Its Management*. Elsevier Science.
- Galetti, M., Pedrosa, F., Keuroghlian, A., Sazima, I., 2016. Liquid lunch – vampire bats feed on invasive feral pigs and other ungulates. *Front. Ecol. Environ.* 14, 505–506. <http://dx.doi.org/10.1002/fee.1431>.
- Galetti, M., Pires, A.S., Brancalion, P.H.S., Fernandez, F.A.S., 2017. Reversing defaunation by trophic rewilding in empty forests. *Biotropica* 49, 5–8. <http://dx.doi.org/10.1111/btp.12407>.
- Genes, L., Fernandez, F.A.S., Vaz-de-Mello, F.Z., da Rosa, P., Fernandez, E., Pires, A.S., 2019. Effects of howler monkey reintroduction on ecological interactions and processes. *Conserv. Biol.* 33, 88–98. <http://dx.doi.org/10.1111/cobi.13188>.
- Gnocchi, A.P., Srbeek-Araujo, A.C., 2017. *Common Vampire Bat (Desmodus rotundus) feeding on Lowland Tapir (Tapirus terrestris) in an Atlantic Forest remnant in southeastern Brazil*. *Biota Neotrop.* 17.
- Gonçalves, F., Fischer, E., Dirzo, R., 2017. *Forest conversion to cattle ranching differentially affects taxonomic and functional groups of Neotropical bats*. *Biol. Conserv.* 210, 343–348.
- Gonçalves, F., Magioli, M., Bovendorp, R.S., Ferraz, K.M.P.M.B., Bulascoschi, L., Moreira, M.Z., Galetti, M., 2020. Prey choice of introduced species by the common vampire bat (*Desmodus rotundus*) on an Atlantic Forest Land-Bridge Island. *Acta Chiropt.* 22, 167–174. <http://dx.doi.org/10.3161/15081109acc2020.22.1.015>.
- Greenhall, A.M., 1972. The biting and feeding habits of the vampire bat, *Desmodus rotundus*. *J. Zool.* 168, 451–461.
- Greenhall, A.M., Joermann, G., Schmidt, U., 1983. *Desmodus rotundus*. *Mamm. Species* 202, 1–6.
- Hernández-Pérez, E.L., Castillo-Vela, G., García-Marmolejo, G., López, M.S., Reyna-Hurtado, R., 2019. Wild pig (*Sus scrofa*) as prey of the common vampire bat (*Desmodus rotundus*). *Therya* 10, 195–199.
- Ibanez, C.J., 1981. *Biología and Ecología de los Murciélagos del Hato “El frío” Apure, Venezuela*. *Acta Vert.* 1, 8–14.
- Johnson, N., Aréchiga-Ceballos, N., Aguilar-Setien, A., 2014. Vampire bat rabies: ecology, epidemiology and control. *Viruses* 6, 1911–1928. <http://dx.doi.org/10.3390/v6051911>.
- Jule, K.R., Leaver, L.A., Lea, S.E.G., 2008. The effects of captive experience on reintroduction survival in carnivores: a review and analysis. *Biol. Conserv.* 141, 355–363. <http://dx.doi.org/10.1016/j.biocon.2007.11.007>.
- Lafferty, K.D., Gerber, L.R., 2002. Good medicine for conservation biology: the intersection of epidemiology and conservation theory. *Conserv. Biol.* 16, 593–604. <http://dx.doi.org/10.1046/j.1523-1739.2002.00446.x>.
- Letko, M., Seifert, S.N., Olival, K.J., Plowright, R.K., Munster, V.J., 2020. Bat-borne virus diversity, spillover and emergence. *Nat. Rev. Microbiol.* 18, 461–471. <http://dx.doi.org/10.1038/s41579-020-0394-z>.
- Lord, R.D., Delpietro, H., Lazaro, L., 1973. *Vampiros que se alimentan de murciélagos*. *Physis Secc.* 32, 225.
- Lovari, S., Ferretti, F., Corazza, M., Minder, I., Troiani, N., Ferrari, C., Saggi, A., 2014. Unexpected consequences of reintroductions: competition between reintroduced red deer and Apennine chamois. *Anim. Conserv.* 17, 359–370. <http://dx.doi.org/10.1111/acv.12103>.
- Luna-Jorquera, G., Culik, B.M., 1995. Penguins bled by vampires. *J für Ornith.* 136, 471–472.
- Mann, G., 1951. *Biología del vampiro*. *Biologica* 12 (13), 3–24.
- McCarthy, T.J., 1989. Human depredation by vampire bats (*Desmodus rotundus*) following a hog cholera campaign. *Am. J. Trop. Med. Hyg.* 40, 320–322. <http://dx.doi.org/10.4269/ajtmh.1989.40.320>.
- Meza, D.K., Broos, A., Becker, D.J., Behdenna, A., Willett, B.J., Viana, M., Streicker, D.G., 2020. Predicting the presence and titre of rabies virus-neutralizing antibodies from low-volume serum samples in low-containment facilities. *Transbound. Emerg. Dis.* n/a. <http://dx.doi.org/10.1111/tbed.13826>.
- Mittelman, P., Kreisler, C., Pires, A.S., Fernandez, F.A.S., 2020. Agouti reintroduction recovers seed dispersal of a large-seeded tropical tree. *Biotropica* 52, 766–774. <http://dx.doi.org/10.1111/btp.12788>.
- Moreno-Mateos, D., Alberdi, A., Morrién, E., van der Putten, W.H., Rodríguez-Uña, A., Montoya, D., 2020. The long-term restoration of ecosystem complexity. *Nat. Ecol. Evol.* 4, 676–685. <http://dx.doi.org/10.1038/s41559-020-1154-1>.
- Nogués-Bravo, D., Simberloff, D., Rahbek, C., Sanders, N.J., 2016. Rewilding is the new Pandora's box in conservation. *Curr. Biol.* 26, R87–R91. <http://dx.doi.org/10.1016/j.cub.2015.12.044>.
- Olival, K.J., Hosseini, P.R., Zambrana-Torrel, C., Ross, N., Bogich, T.L., Daszak, P., 2017. Host and viral traits predict zoonotic spillover from mammals. *Nature* 546, 646–650. <http://dx.doi.org/10.1038/nature22975>.
- Perino, A., Pereira, H.M., Navarro, L.M., Fernández, N., Bullock, J.M., Ceausu, S., Cortés-Avizanda, A., van Klink, R., Kuemmerle, T., Lomba, A., Petetquotrighter, G., Plieninger, T., Rey Benayas, J.M., Sandom, C.J., Svenning, J.-C., Wheeler, H.C., 2019. Rewilding complex ecosystems. *Science* (80-.) 364. <http://dx.doi.org/10.1126/science.aav5570>.
- Pires, M.M., 2017. Rewilding ecological communities and rewiring ecological networks. *Perspect. Ecol. Conserv.* 15, 257–265. <http://dx.doi.org/10.1016/j.pecon.2017.09.003>.
- Recuenco, S.E., 2020. Rabies vaccines, prophylactic, Peru: massive rabies pre-exposure prophylaxis for High-risk populations. In: Ertl, H.C.J. (Ed.), *Rabies and Rabies Vaccines*. Springer International Publishing, Cham, pp. 83–101. http://dx.doi.org/10.1007/978-3-030-21084-7_5.
- Rodríguez-San Pedro, A., Allendes, J.L., 2017. Echolocation calls of free-flying common vampire bats *Desmodus rotundus* (Chiroptera: phyllostomidae) in Chile. *Bioacoustics* 26, 153–160. <http://dx.doi.org/10.1080/09524622.2016.1231079>.
- Sánchez-Cordero, V., Botello, F., Magaña-Cota, G., Iglesias, J., 2011. *Vampire bats, Desmodus rotundus, feeding on white-tailed deer, Odocoileus virginianus*. *Mammalia* 75 (1), 91–92.
- Serrano, H., Pérez-Rivero, J.-J., Aguilar-Setián, A., de-Paz, O., Villa-Godoy, A., 2007. *Vampire bat reproductive control by a naturally occurring phytoestrogen*. *Reprod. Fertil. Dev.* 19, 470–472.
- Stoner-Duncan, B., Streicker, D.G., Tedeschi, C.M., 2014. Vampire bats and rabies: toward an ecological solution to a public health problem. *PLoS Negl. Trop. Dis.* 8, 1–5. <http://dx.doi.org/10.1371/journal.pntd.0002867>.
- Streicker, D.G., Allgeier, J.E., 2016. Foraging choices of vampire bats in diverse landscapes: potential implications for land-use change and disease transmission. *J. Appl. Ecol.* 53, 1280–1288. <http://dx.doi.org/10.1111/1365-2664.12690>.
- Streicker, D.G., Turmelle, A.S., Vonhof, M.J., Kuzmin, I.V., McCracken, G.F., Rupprecht, C.E., 2010. Host phylogeny constrains cross-species emergence and establishment of rabies virus in bats. *Science* (80-.) 329, 676–679. <http://dx.doi.org/10.1126/science.1188836>.
- Streicker, D.G., Recuenco, S., Valderrama, W., Benavides, J.G., Vargas, I., Pacheco, V., Condori Condori, R.E., Montgomery, J., Rupprecht, C.E., Rohani, P., Altizer, S.,

2012. Ecological and anthropogenic drivers of rabies exposure in vampire bats: implications for transmission and control. *Proc. R. Soc. B Biol. Sci.* 279, 3384–3392, <http://dx.doi.org/10.1098/rspb.2012.0538>.
- Svenning, J.-C., Pedersen, P.B.M., Donlan, C.J., Ejrnæs, R., Faurby, S., Galetti, M., Hansen, D.M., Sandel, B., Sandom, C.J., Terborgh, J.W., Vera, F.W.M., 2016. Science for a wilder Anthropocene: synthesis and future directions for trophic rewilding research. *Proc. Natl. Acad. Sci.* 113, 898–906, <http://dx.doi.org/10.1073/pnas.1502556112>.
- Thompson, R.D., Mitchell, G.C., Burns, R.J., 1972. Vampire bat control by systemic treatment of livestock with an anticoagulant. *Science*. 177, 806–808, <http://dx.doi.org/10.1126/science.177.4051.806>.
- United Nations, Available from http://www.un.org/en/ga/search/view_doc.asp?symbol=A/RES/73/284 (accessed August 2020) 2019. United Nations Decade on Ecosystem Restoration (2021–2030).
- Villa, R.B., Lopez-Forment, W., 1966. Cinco casos de predacion de pequenos vertebrados en murciélagos de Mexico. *An. Inst. Biol. Univ. Nac. Auton. Mex. Ser. Zool.* 40 (2), 291–298.
- Voigt, C.C., Kelm, D.H., 2006. Host preference of the common vampire bat (*Desmodus rotundus*; Chiroptera) Assessed by stable isotopes. *J. Mammal.* 87, 1–6, <http://dx.doi.org/10.1644/05-MAMM-F-276R1.1>.
- Zortéa, M., Silva, D.A., Calaça, A.M., 2018. Susceptibility of targets to the vampire bat *Desmodus rotundus* are proportional to their abundance in Atlantic Forest fragments? *Iheringia. Série Zool.* 108, <http://dx.doi.org/10.1590/1678-4766e2018037>.